

What controls microbial enzyme activity in wetlands?

Colin R. Jackson

Department of Biology, University of Mississippi

cjackson@olemiss.edu

Everything

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities...were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after sewage effluent addition (Simpson et al. 2002)

Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate than in the bottom layer (Simpson et al. 2002)

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were not affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with increasing salinity (Simpson et al. 2002)

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al. 2008)

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single factor (Menon et al. 2013)

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2 efflux, extractable DOC, simple substrate utilization, and microbial biomass (Kang and Freeman 1999)

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulase and acid phosphatase were strongly depended on plant species richness (Zhang et al. 2008)

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kang and Freeman 1999)

All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al. 2008)

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to soil moisture content (Sinsabaugh et al. 2008)

Enzyme activity decreased with depth and showed significant variation over the growing season. Site-specific factors such as nutrient availability explain deviations (Pinsonneault et al. 2016)

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities... were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulase and acid phosphatase were strongly depended on plant species richness (Zhang et al.

Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate than

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al.

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kan

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2

All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al.

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single

efflux, extractable DOC, simple substrates and

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

Enzyme activity decreased with depth and showed significant variation over the growing season. Site-specific factors such as nutrient availability explain deviations (Pinsonneault et al. 2016)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities... were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after

Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate that

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were not affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al.

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2 efflux, extractable DOC, simple

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulase and acid phosphatase were strongly depended on plant species richness (Zhang et al.

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kang

All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al.

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to

Enzyme activity decreased with depth and showed significant variation over the growing season. Site-specific factors such as nutrient availability explain deviations (Pinsonneault et al. 2016)

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities... were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after

sewage effluent addition (Jackson and Vallaire 2009). Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate that

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al. 2007)

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2 efflux, extractable DOC, simple

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulase and acid phosphatase were strongly depended on plant species richness (Zhang et al. 2016)

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kang and Freeman 1999)

All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al. 2008)

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to

Enzyme activity decreased with depth and showed significant variation over the growing season. Site-specific factors such as nutrient availability explain deviations (Pinsonneault et al. 2016)

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities... were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after

Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate that

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al.)

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2 efflux, extractable DOC, simple

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulase and acid phosphatase were strongly depended on plant species richness (Zhang et al.)

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kang and Freeman 1999)

All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al.)

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to

Enzyme activity decreased with depth and showed significant variation over the growing season. Site-specific factors such as nutrient availability explain deviations (Pinsonneault et al. 2016)

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities... were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after

sew 200 Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate that

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al.

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2 efflux, extractable DOC, simple

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulase and acid phosphatase were strongly depended on plant species richness (Zhang et al.

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kang

All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al.

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to

Enzyme activity decreased with depth and showed significant variation over the growing season. Site-specific factors such as nutrient availability explain deviations (Pinsonneault et al. 2016)

..compared to controls receiving no carbon, phosphatase showed no significant differences. Sulphatase activities... were significantly reduced by glucose, cellulose and sewage effluent addition. β -glucosidase activity gradually increased in response to cellulose addition, and decreased after

Activities of all enzymes were significantly correlated with root activity in *Vetiveria zizanioides* and *Phragmites australis* wetlands, but not in *Hymenocallis littoralis* wetlands. Significant correlations between enzyme activity, root biomass and root growth were found in *Cyperus flabelliformis* wetlands. Activities of phosphatase and cellulase were high in the top layer of the substrate that

Activities of β -glucosidase, chitinase and phosphatase differed widely among species but were poorly related to litter nutrient concentrations. Within some species, phosphatase activity increased towards high litter N:P ratios (Güsewell and Freeman 2005)

Exposure to elevated salinity also decreased phosphatase and NAGase activity by almost 20%, with less effect on β -glucosidase. P addition had no impact on extracellular enzyme activity. (Jackson and Vallaire 2009)

Alkaline phosphatase activity was affected by P loading and was negatively related to soil P concentrations and microbial biomass and P. Arylsulfatase, β -d-glucosidase, protease, and phenol oxidase were affected by P loading and were not related to measured soil C, N, S, physical and chemical parameters. Enzyme activities decreased with

Enzyme activity was correlated with sediment and water chemistry and stoichiometry, N deposition, the agricultural stress gradient and hydrological turnover time (Hill et al.

β -1,4-glucosidase, phosphatase, and NAGase exhibited similar activity for all vegetation treatments, while the activity of phenol oxidase and peroxidase was higher in sediments with no vegetation (Menon et al. 2013)

Overall, NAGase were the lowest in bogs and much higher in freshwater marshes and flooded grasslands. The variations of the activity were not explained by a single

O_2 availability and the activities of some enzymes appeared to be related at landscape scales after accounting for differences in organic matter. Reducing conditions and phenolic compounds did not appear to constrain soil hydrolytic enzyme activity (Hall et al. 2014)

Phosphatase activity was suppressed in P-addition plots under all salinity levels while activities of the remaining enzymes were higher in P-enriched plots (Reimánková and Sirová 2007)

Hydrogen ion concentration was a dominant controlling factor for the phosphatase activities. Waterlogging and low temperature seem to restrict enzyme activities in fen and swamp sites, as both factors showed correlations with enzyme activities. A negative relationship between phosphatase activity and phosphate content was discernable, when compared on a spatial basis. (Kang and Freeman 1999)

Decreases in activities of β -glucosidase, NAGase, phosphatase, and phenol oxidase, and soil pH were observed with H_4NO_3 . Under alkaline conditions, marginal changes in response to N additions were observed in the soil CO_2

Enzyme activities were different among vegetation types. Soil MBC content was significantly correlated with activities of β -glucosidase. DOC content was significantly correlated with the activities of alkaline phosphomonoesterase (Shao et al. 2015)

pH, temperature, soluble phenolics, total organic carbon, phosphorus, and nitrogen significantly influenced enzyme activities (Luo and Gu 2014)

Activities of β -glucosidase, phenol oxidase, protease and nitrate reductase, while affected by plant species richness, were strongly depended on the presence or absence of plants. Activities of cellulose and acid phosphatase were strongly depended on plant species richness (Zhang et al.

Elevated CO_2 had no effect on β -glucosidase activity. However, NAGase activity increased significantly in cores from the bog, whilst a similar response was found in the gully mire for phosphatase. Such changes were absent from the fen and marsh where inorganic nutrients were abundant, suggesting that enzyme activities involved in N or P mineralisation only increase under elevated CO_2 when nutrient limitation is strongly exerted. (Kang

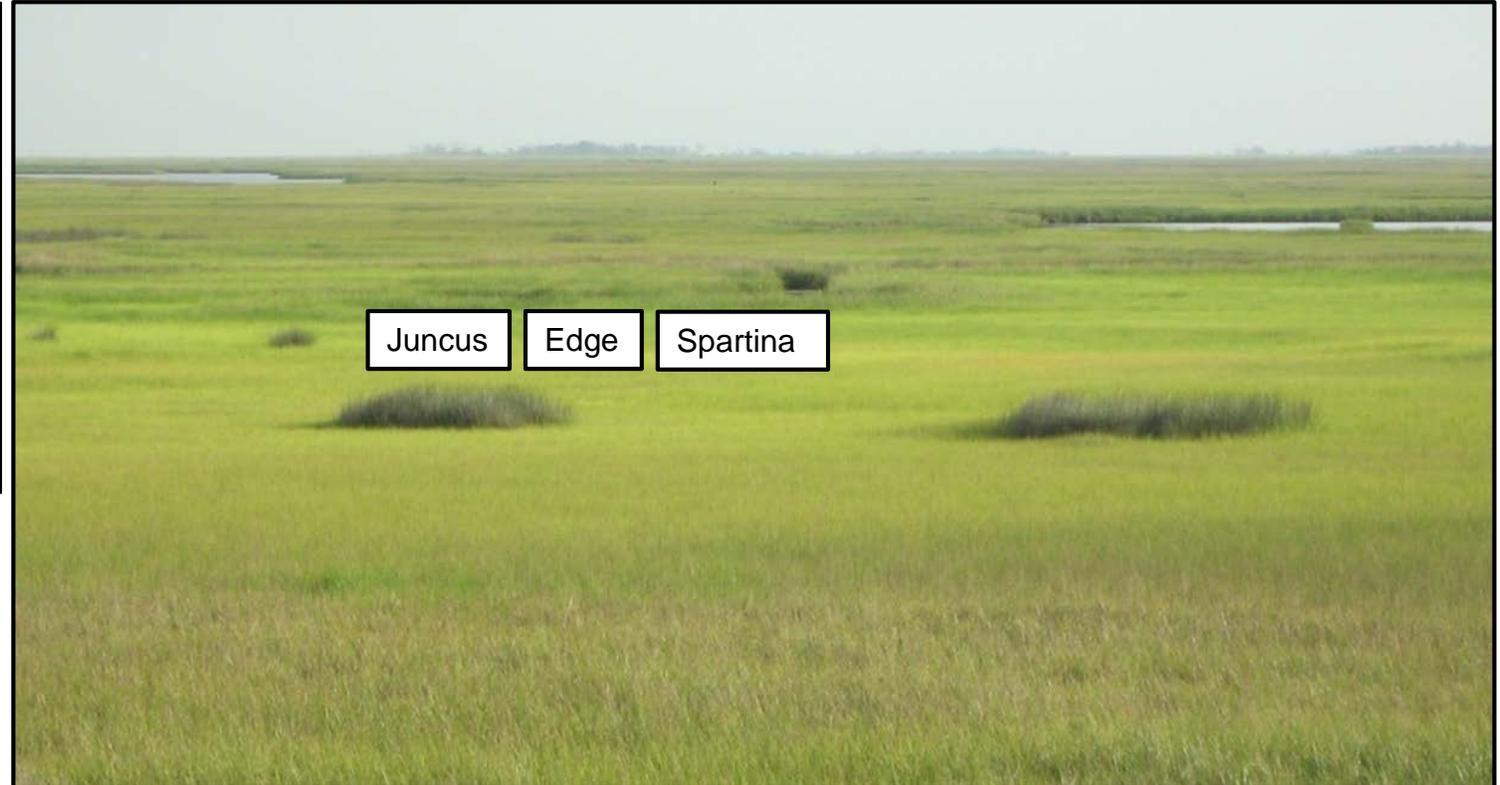
All activities were significantly related to soil pH. Oxidative activities were more variable than hydrolytic activities and increased with soil pH (Sinsabaugh et al.

β -glucosidase, NAGase, and phosphatase were stimulated under drying condition. Increase of enzyme activities under drying was related to

Enzyme activity decreased with depth and showed significant variation over growing season as nutrient availability explain deviations (Pinsonneault et al. 2016)

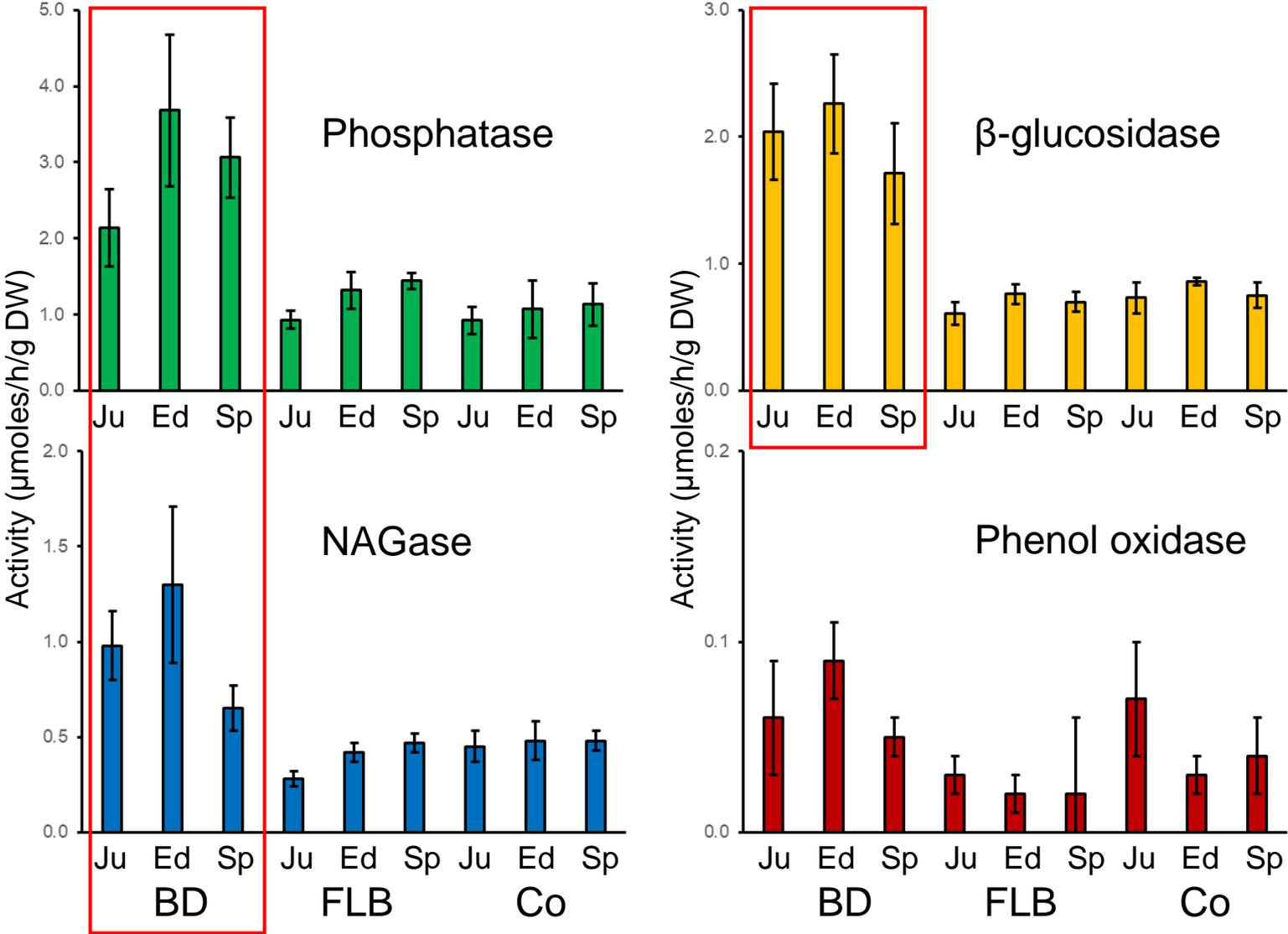
Site-specific factors

Does wetland vegetation type control microbial enzyme activity in Gulf Coast wetlands?



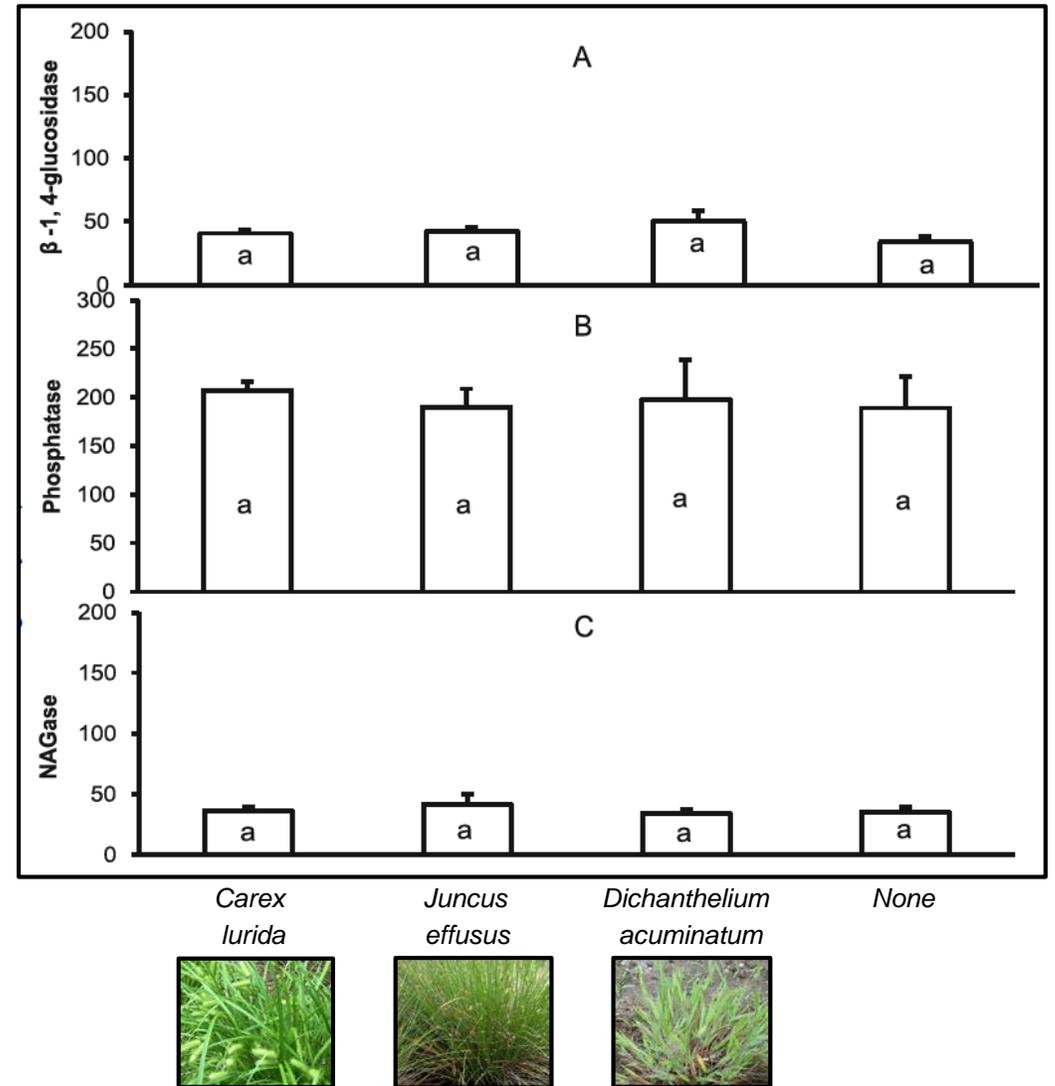
Rietl et al. 2016. Microbial community composition and extracellular enzyme activity associated with *Juncus roemerianus* and *Spartina alterniflora* vegetated sediments in Louisiana saltmarshes. *Microbial Ecology* 71:290-303.

Enzyme activity was influenced by site more than vegetation type



Rietl et al. 2016. Microbial community composition and extracellular enzyme activity associated with *Juncus roemerianus* and *Spartina alterniflora* vegetated sediments in Louisiana saltmarshes. *Microbial Ecology* 71:290-303.

Vegetation did not influence the activity of β -glucosidase, phosphatase, or NAGase in wetland mesocosms



Why is it so hard to determine what controls enzyme activity in wetlands?

Carbon?

Vegetation?

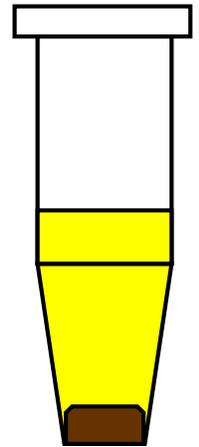
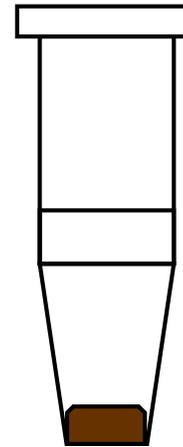
Abiotic factors?

Nutrients?

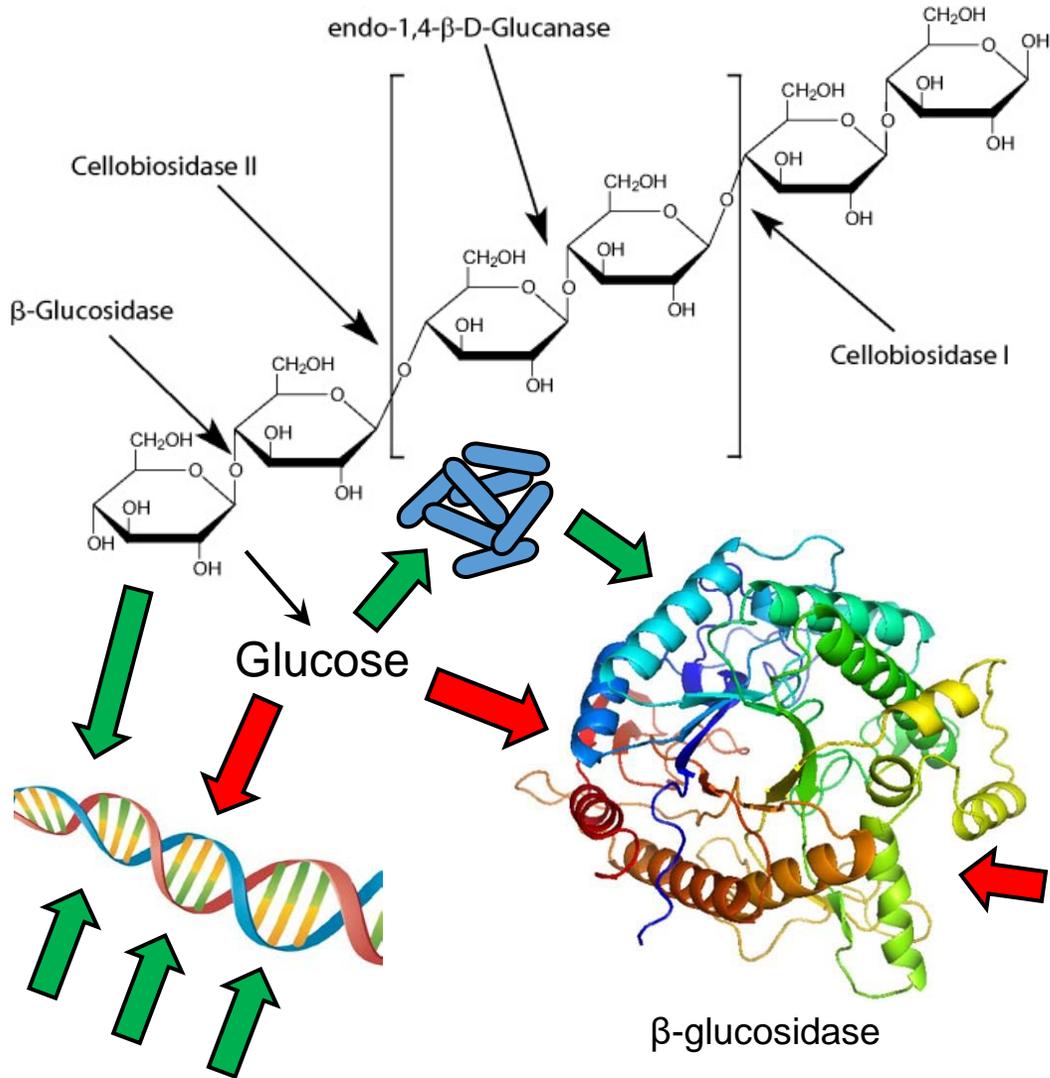
Microorganisms?

Site-specific?

Analysis of enzymes in wetlands focuses at an ecological level



How does carbon affect β -glucosidase activity?



Glucose

Inhibition

Repression

Enzyme concentration

Cellulose/cellobiose

Induction

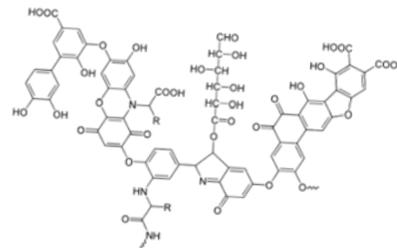
Other substrates

Induction

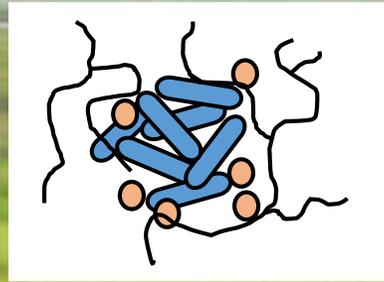
Phenolics

Inhibition

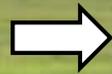
So how does carbon affect β -glucosidase activity?



Analysis of enzymes in wetlands focuses at an ecological level



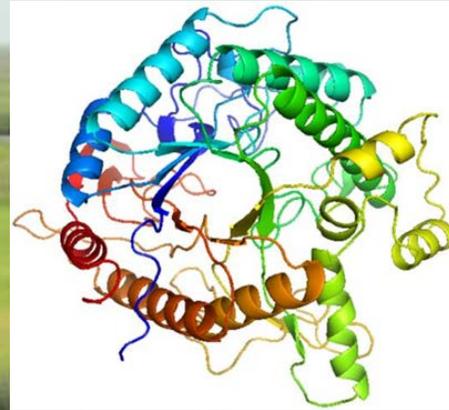
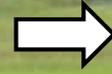
Microbiome



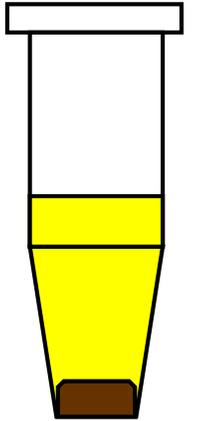
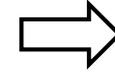
Genome
(metagenome)



Transcriptome

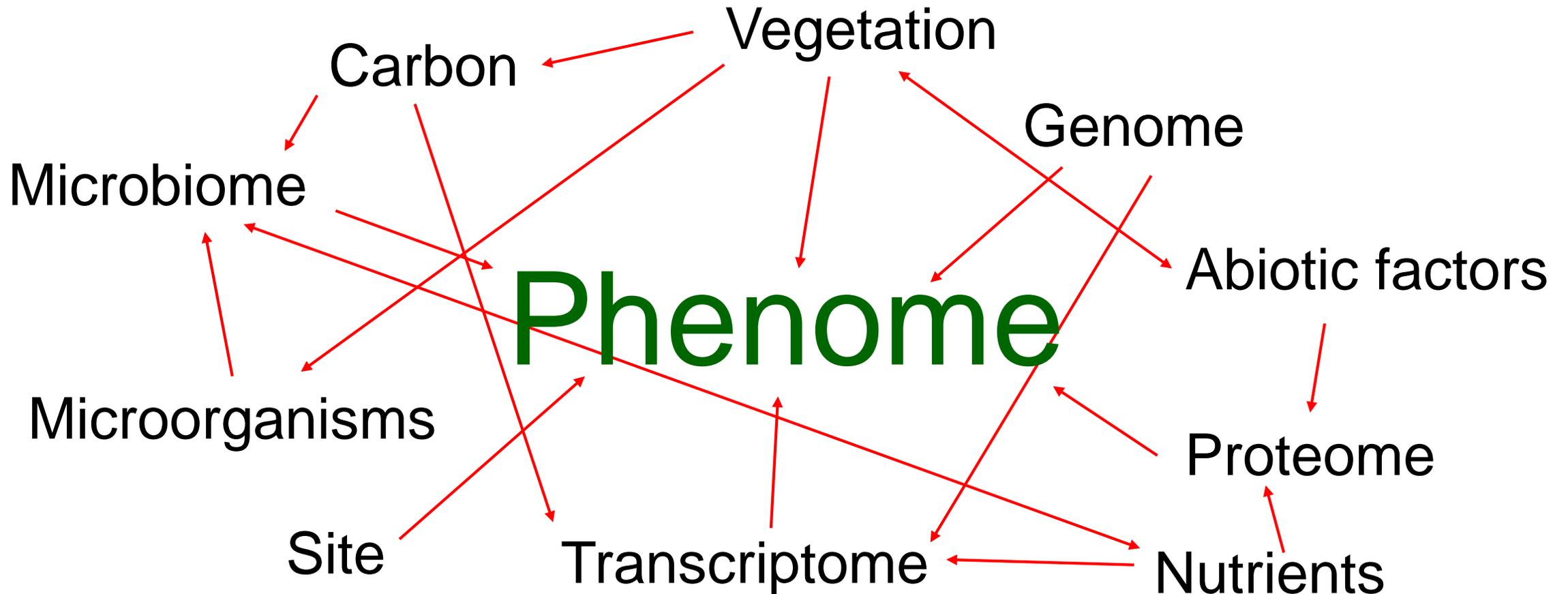


Proteome



Phenome

The **phenome** is the phenotypes expressed by a cell, tissue, organ, organism, or species (**ecosystem?**)



Acknowledgements

Former students and collaborators:

Anthony Rietl
Rani Menon

Current students:

Eric Weingarten
Bram Stone
Sarah Russell

